



ORA™ SEE aPCR Probe Mix, 2X

CAT.#	SIZE	COMPONENTS	COMPONENT COMPOSITION
ODD0401	200 r of	2 x 1 ml - ORA™ SEE qPCR Probe Mix, 2X	Mix includes an inert blue dye for better visibility, Hot Start qPCR
QPP0401	20 μΙ	2 x 1 ml - PCR Water	components: dNTPs at 0.25 mM, optimized buffer; ROX is not included.
	1000 r of	10 x 1 ml - ORA™ SEE qPCR Probe Mix, 2X	Mix includes an inert blue dye for better visibility, Hot Start qPCR
QPP0405	20 μΙ	10 x 1 ml - PCR Water	components: dNTPs at 0.25 mM, optimized buffer; ROX is not included.
Storage:	In the dark at -20°C.		

APPLICATIONS

- qPCR assays based on specific probes: including TagMan[®], Molecular Beacons, Scorpions™ Probes
- Quantification of gDNA, cDNA, viral DNA, low copy number genes, gene expression analysis

PRODUCT DETAILS

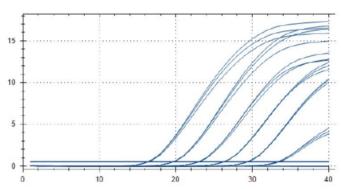
highQu qPCR mastermixes are based on the small molecular inhibitor technology Hot Start PCR allowing to achieve highest sensitivity and specificity under both standard and fast qPCR cycling conditions. They provide excellent results on both AT and GC rich templates, in multiplexing and guaranty rapid extension with early Ct values with minimum or no optimization. ORA™ SEE qPCR mixes provide an additional advantage of a simplified tracking of the process, as they are colored with an inert blue dye to make samples much better visible during pipetting and handling.

Our mastermixes are supplied with PCR Water to guaranty the best performance. To suit the broad instrument range the ORA™ qPCR Probe Mixes are available in three versions - without ROX, with low or high ROX concentration.

BENEFITS

- Universal both standard and fast cycling, all probe qPCR assays, GC or AT rich templates
- · Excellent for both single-plex & multiplexing
- Rapid extension, early Ct
- Inert blue dye for a better sample visibility and tracking

PERFORMANCE



Visible blue samples, 100% efficiency, 10 copies detection sensitivity achieved with ORA™ SEE qPCR Probe Mixes. TagMan® probe amplification with ORA™ SEE qPCR Probe Mix from plasmid dilution series (1 x 10⁶ to 10 copies).



PROTOCOL

- Use special primer selection programs for good planning.
- Work with amplicons in a range of 80-200, max 400 bp.
- Take typical measures to prevent PCR cross over contamination, keep your bench clean, wear gloves, use sterile tubes and filter pipet tips.
- Run reactions in triplets; include a no-template control and positive control in parallel.
- Thaw and keep reagents on ice. Mix well before use.
- Do not perform annealing/extension for more than 30 seconds and do not use lower than 60 °C temperature for this step.

✓	Prepare a 20 µl reaction:
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Reverse Primer	100-400 nM final c.
Forward Primer	100-400 nM final c.
Specific Probe	200 nM final c. (0.4 μl of 10 μM)
cDNA Template or	<100 ng <i>or</i>
gDNA Template	1 μg
PCR Water	to 10 μl
ORA™ SEE qPCR Mix, 2X	10 μl

- Mix gently, avoid bubbles.
- Place into the instrument set like:

Initial denaturation	1 cycle: 95°C - 2 min for cDNA, or
	1 cycle: 95°C - 3 min for gDNA
Denaturation	40 cycles: 95°C - 5 sec
Anneal./extension	40 cycles: 60-65°C – 20-30 sec

✓ Follow instrument instructions for melting curve analysis.

IN VITRO RESEARCH USE ONLY